	٩
Oregon State Micronutrient	Information Center
MIC Home About 🔽 Articles 🔽 Español	Resources 🔽 Disease Index Giving LPI Home
Home » Dietary Factors » Phytochemic	als
The Micronutrient Information Center is please help by <u>donating to the LPI Outro</u>	supported by your donations. If you value this website, each Education Fund.
Vitamins	
Minerals	
Other Nutrients	
Dietary Factors	
Food and Beverages	
Micronutrients and Health	
Life Stages	
Flavonoids	
Contonto	Summary
Contents <ul> <li>Summary</li> <li>Introduction</li> </ul>	<ul> <li>Flavonoids are a large family of polyphenolic compounds synthesized by plants. <u>(More</u> <u>information)</u></li> </ul>

- Scientists are interested in the potential health benefits of flavonoids associated with fruit and vegetable-rich diets. <u>(More information)</u>
- Many of the biological effects of flavonoids appear to be related to their ability to modulate cellsignaling pathways, rather than their antioxidant activity. <u>(More information)</u>
- Although higher intakes of flavonoid-rich foods are associated with reductions in cardiovascular disease

Disease Prevention

Biological Activities

<u>Cardiovascular disease</u>
 <u>Cancer</u>

Metal chelation

Metabolism and Bioavailability

Direct antioxidant activity

> Effects on cell signaling

- <u>Neurodegenerative diseases</u>
- <u>Sources</u>
  - Food
  - > Supplements
- Safety
  - Adverse effects
  - Pregnancy and lactation
  - Drug interactions
  - Nutrient interactions
- Authors and Reviewers
- References

risk, it is not yet known whether flavonoids themselves are cardioprotective. <u>(More information)</u>

- Despite promising results in animal studies, it is not clear whether high flavonoid intakes can help prevent cancer in humans. <u>(More information)</u>
- It is not yet clear how flavonoid consumption affects neurodegenerative disease risk in humans. <u>(More information)</u>
- Higher intakes of flavonoid-rich foods have been associated with reduced risk of chronic disease in some studies, but it is not known whether isolated flavonoid supplements or extracts will confer the same benefits as flavonoid-rich foods.

### Introduction

Flavonoids are a large family of compounds synthesized by plants that have a common chemical structure (**Figure 1**). Flavonoids may be further divided into subclasses (**Table 1**). Over the past decade, scientists have become increasingly interested in the potential for various dietary flavonoids to explain some of the health benefits associated with fruit- and vegetable-rich diets. This article reviews the scientific evidence for the hypothesis that dietary flavonoids promote health and prevent disease in humans. For more detailed information on the health effects of isoflavones, a subclass of flavonoids with estrogenic activity, see the separate article on <u>Soy</u> <u>Isoflavones</u>. For more information on the health benefits of foods that are rich in flavonoids, see the separate articles on <u>Fruit and Vegetables</u>, <u>Legumes</u>, and <u>Tea</u>.

#### Figure 1. Basic Chemical Structure of a Flavonoid



### Table 1. Common Dietary Flavonoids (Select the highlighted text to view chemical structures.)

Flavonoid Subclass	Dietary Flavonoids	Some Common Food Sources
Anthocyanidins	Cyanidin, Delphinidin, Malvidin, Pelargonidin, Peonidin, Petunidin	Red, blue, and purple berries; red and purple grapes; red wine
Flavanols	Monomers (Catechins): Catechin, Epicatechin, Epigallocatechin Epicatechin gallate, Epigallocatechin gallate Dimers and Polymers:	<b>Catechins</b> : Teas (particularly green and white), chocolate, grapes, berries, apples <b>Theaflavins, Thearubigins</b> : Teas

	<u>Theaflavins</u> , Thearubigins, <u>Proanthocyanidins</u>	(particularly black and oolong) <b>Proanthocyanidins</b> : Chocolate, apples, berries, red grapes, red wine
Flavanones	Hesperetin, Naringenin, Eriodictyol	Citrus fruit and juices, e.g., oranges, grapefruit, lemons
Flavonols	Quercetin, Kaempferol, Myricetin, Isorhamnetin	Widely distributed: yellow onions, scallions, kale, broccoli, apples, berries, teas
<b>Flavones</b>	Apigenin, Luteolin	Parsley, thyme, celery, hot peppers,
<b>Isoflavones</b>	Daidzein, Genistein, Glycitein	Soybeans, soy foods, legumes

### Figure 2. Chemical Structures of Some Anthocyanidins (Anthocyanin Aglycones)





## Figure 4. Chemical Structures of Theaflavins (Flavanol Dimers)



http://lpi.oregonstate.edu/mic/dietary-factors/phytochemicals/flavonoids[2016/01/28 21:36:31]







### **Figure 6. Chemical Structures of Some Flavanones**







## Metabolism and Bioavailability

Flavonoids connected to one or more sugar molecules are known as flavonoid <u>alvcosides</u>, while those that are not connected to a sugar molecule are called aglycones. With the exception of flavanols (catechins and proanthocyanidins), flavonoids occur in plants and most foods as glycosides (2). Even after cooking, most flavonoid glycosides reach the small intestine intact. Only flavonoid aglycones and flavonoid glucosides (bound to glucose) are absorbed in the small intestine, where they are rapidly metabolized to form methylated, glucuronidated, or sulfated <u>metabolites (3)</u>. Bacteria that normally colonize the <u>colon</u> also play an important role in flavonoid metabolism and absorption. Flavonoids or flavonoids metabolites that reach the colon may be further metabolized by bacterial enzymes, and then absorbed. A person's ability to produce specific flavonoid metabolites may vary and depends on the milieu of the colonic microflora (4, 5). In general, the <u>bioavailability</u> of flavonoids is relatively low due to limited absorption and rapid elimination. Bioavailability differs for the various flavonoids: isoflavones are the most bioavailable group of flavonoids, while flavanols (proanthocyanidins and tea catechins) and anthocyanins are very poorly absorbed (6). Since flavonoids are rapidly and extensively metabolized, the biological activities of flavonoid metabolites are not always the same as those of the parent compound (reviewed in  $\underline{7}$ ) When evaluating the data from flavonoid research in cultured cells, it is important to consider whether the flavonoid concentrations and metabolites used are physiologically relevant (8). In humans, peak <u>plasma</u> concentrations of soy isoflavones and citrus flavanones have not been found to exceed 10 micromoles/liter after oral consumption. Peak plasma concentrations measured after the consumption of anthocyanins, flavanols and flavonols (including those from tea) are generally less than 1 micromole/liter (3).

## **Biological Activities**

### **Direct antioxidant activity**

Flavonoids are effective scavengers of free radicals in the test tube (*in vitro*) (9, 10). However, even with very high flavonoid intakes, plasma and intracellular flavonoid concentrations in humans are likely to be 100-1,000 times lower than concentrations of other <u>antioxidants</u>, such as ascorbate (vitamin C), uric acid, or glutathione. Moreover, most circulating flavonoids are actually flavonoid <u>metabolites</u>, some of which have lower antioxidant activity than the parent flavonoid. For these reasons, the relative contribution of dietary flavonoids to plasma and tissue antioxidant function *in vivo* is likely to be very small or negligible (7, 11, 12).

## **Metal chelation**

Metal ions, such as iron and copper, can <u>catalyze</u> the production of <u>free radicals</u>. The ability of flavonoids to chelate (bind) metal ions appears to contribute to their antioxidant activity <u>in vitro</u> (<u>13, 14</u>). In living organisms, most iron and copper are bound to proteins, limiting their participation in reactions that produce free radicals. Although the metal-chelating activities of flavonoids may be beneficial in pathological conditions of iron or copper excess, it is not known whether flavonoids or their metabolites function as effective metal chelators <u>in vivo</u> (<u>11</u>).

### Effects on cell-signaling pathways

Cells are capable of responding to a variety of different stresses or signals by increasing or decreasing the availability of specific proteins. The complex cascades of events that lead to changes in the expression of specific genes are known as <u>cell-signaling</u> pathways or signal transduction pathways. These pathways regulate numerous cell processes, including growth, proliferation, and death (apoptosis). Although it was initially hypothesized that the biological effects of flavonoids would be related to their antioxidant activity, available evidence from cell culture experiments suggests that many of the biological effects of flavonoids are related to their ability to modulate cell-signaling pathways (7). Intracellular concentrations of flavonoids required to affect cell-signaling pathways are considerably lower than those required to affect cellular antioxidant capacity. Flavonoid metabolites may retain their ability to interact with cell-signaling proteins even if their antioxidant activity is diminished (15, 16). Effective signal transduction requires proteins known as kinases that <u>catalyze</u> the <u>phosphorylation</u> of target proteins at specific sites. Cascades involving specific phosphorylations or dephosphorylations of signal transduction proteins ultimately affect the activity of transcription factors - proteins that bind to specific response elements on DNA and promote or inhibit the transcription of various genes. The results of numerous studies in cell culture suggest that flavonoids may affect chronic disease by selectively inhibiting kinases (7, 17). Cell growth and proliferation are also regulated by growth factors that initiate cell-signaling cascades by binding to specific receptors in cell membranes. Flavonoids may alter growth factor signaling by inhibiting receptor phosphorylation or blocking receptor binding by growth factors (18).

Modulation of cell-signaling pathways by flavonoids could help prevent cancer by:

**Stimulating phase II detoxification enzyme activity** (<u>19, 20</u>): Phase II detoxification enzymes <u>catalyze</u> reactions that promote the <u>excretion</u> of potentially toxic or <u>carcinogenic</u> chemicals.

**Preserving normal cell cycle regulation** (21, 22): Once a cell divides, it passes through a sequence of stages collectively known as the cell cycle before it divides again. Following DNA damage, the cell cycle can be transiently arrested at damage checkpoints, which allows for DNA repair or activation of pathways leading to cell death (apoptosis) if the damage is irreparable (23). Defective cell cycle regulation may result in the propagation of mutations that contribute to the development of cancer.

**Inhibiting proliferation and inducing apoptosis** (24-26): Unlike normal cells, cancer cells proliferate rapidly and lose the ability to respond to cell death signals that initiate apoptosis.

**Inhibiting tumor invasion and angiogenesis** (27, 28): Cancerous cells invade normal tissue aided by <u>enzymes</u> called matrix-metalloproteinases. To fuel their rapid growth, invasive tumors must develop new blood vessels by a process known as angiogenesis.

**Decreasing inflammation** (29-31): Inflammation can result in locally increased production of <u>free radicals</u> by inflammatory enzymes, as well as the release of inflammatory mediators that promote cell proliferation and angiogenesis and inhibit apoptosis (32).

Modulation of cell-signaling pathways by flavonoids could help prevent cardiovascular disease by:

**Decreasing inflammation** (29-31): <u>Atherosclerosis</u> is now recognized as an inflammatory

disease, and several measures of <u>inflammation</u> are associated with increased risk of <u>myocardial</u> <u>infarction</u> (heart attack) (33).

**Decreasing vascular cell adhesion molecule expression** (34, 35): One of the earliest events in the development of atherosclerosis is the recruitment of inflammatory white blood cells from the blood to the arterial wall. This event is dependent on the expression of adhesion molecules by the <u>vascular endothelial</u> cells that line the inner walls of blood vessels (36).

**Increasing endothelial nitric oxide synthase (eNOS) activity** (<u>37</u>): eNOS is the enzyme that catalyzes the formation of nitric oxide by vascular endothelial cells. Nitric oxide is needed to maintain arterial relaxation (<u>vasodilation</u>). Impaired nitric oxide-dependent vasodilation is associated with increased risk of <u>cardiovascular disease</u> (<u>38</u>).

**Decreasing platelet aggregation** (39, 40): <u>Platelet</u> aggregation is one of the first steps in the formation of a blood clot that can occlude a coronary or cerebral artery, resulting in myocardial infarction or stroke, respectively. Inhibiting platelet aggregation is considered an important strategy in the primary and secondary prevention of cardiovascular disease (41).

# **Disease Prevention**

## Cardiovascular disease

### Epidemiological evidence

Several prospective cohort studies conducted in the US and Europe have examined the relationship between some measure of dietary flavonoid intake and coronary heart disease (CHD) risk (42-49). Some studies have found that higher flavonoid intakes to be associated with significant reductions in CHD risk (42-46, 50), while others have reported no significant relationship (47-49, 51). In general, the foods that contributed most to total flavonoid intake in these cohorts were black tea, apples, and onions. One study in the Netherlands also found cocoa to be a significant source of dietary flavonoid intake and the risk of stroke, only two studies found that higher flavonoid intakes were associated with significant reductions in the risk of stroke (45, 52), while five found no relationship (46, 49, 50, 53, 54). Although data from prospective cohort studies suggest that higher intakes of flavonoid-rich foods may help protect against CHD, it cannot be determined whether such protection is conferred by flavonoids, other nutrients and phytochemicals in flavonoid-rich foods, or the whole foods themselves (55).

### Vascular endothelial function

<u>Vascular endothelial</u> cells play an important role in maintaining cardiovascular health by producing nitric oxide, a compound that promotes arterial relaxation (vasodilation) (56). Arterial vasodilation resulting from endothelial production of nitric oxide is termed endothelium-dependent vasodilation. Several clinical trials have examined the effect of flavonoid-rich foods and beverages on endothelium-dependent vasodilation. Two controlled clinical trials found that daily consumption of 4-5 cups (900-1,250 mL) of black tea for four weeks significantly improved endotheliumdependent vasodilation in patients with coronary artery disease (57) and in patients with mildly elevated serum <u>cholesterol</u> levels (58) compared with the equivalent amount of caffeine alone or hot water. Other small clinical trials found similar improvements in endothelium-dependent vasodilation in response to daily consumption of about 3 cups (640 mL) of purple grape juice (59) or a high-flavonoid dark chocolate bar for two weeks (60). More recently, a six-week cocoa intervention trial in 32 postmenopausal women with high cholesterol levels found significant improvements in endothelial function with daily cocoa supplementation (61). Improvements in endothelial function were also noted in conventionally medicated type 2 diabetics following flavanol-rich cocoa supplementation for 30 days (62). The flavanol epicatechin appears to be one of the compounds in flavanol-rich cocoa responsible for its vasodilatory effects (63). Interestingly, a recent randomized controlled trial in 44 older adults found that low doses of flavonoid-rich dark chocolate (6.3 grams/day for 18 weeks; equivalent to 30 calories) increased levels of plasma S-

nitrosoglutathione, an indicator of nitric oxide production, compared to flavonoid-devoid white chocolate (64).

Endothelial nitric oxide production also inhibits the adhesion and aggregation of <u>platelets</u>, one of the first steps in blood clot formation (56). A number of clinical trials have examined the potential for high flavonoid intakes to decrease various measures of platelet aggregation outside of the body (*ex vivo*); such trials have reported mixed results. In general, increasing flavonoid intakes by increasing fruit and/or vegetable intake did not significantly affect ex vivo platelet aggregation (41, 65, 66), nor did increasing black tea consumption (67, 68). However, several small clinical trials in healthy adults have reported significant decreases in ex vivo measures of platelet aggregation after consumption of grape juice (~500 mL/day) for 7-14 days (69-71). Similar inhibition of platelet aggregation has been reported following acute or short-term consumption of dark chocolate (72) and following acute consumption of a flavonoid-rich cocoa beverage (73, 74). In addition, a <u>placebo</u>-controlled trial in 32 healthy adults found that four-week supplementation with flavanols and procyanidins from cocoa inhibited platelet aggregation and function (75). The results of some controlled clinical trials suggest that relatively high intakes of some flavonoid-rich foods and beverages, including black tea, purple grape juice, and cocoa, may improve vascular endothelial function, but it is not known whether these short-term improvements will result in long-term reductions in cardiovascular disease risk.

### Cancer

Although various flavonoids have been found to inhibit the development of chemically-induced cancers in animal models of lung (76), oral (77), esophageal (78), stomach (79), colon (80), skin (81), prostate (82, 83), and mammary (breast) cancer (84), epidemiological studies do not provide convincing evidence that high intakes of dietary flavonoids are associated with substantial reductions in human cancer risk. Most prospective cohort studies that have assessed dietary flavonoid intake using food frequency questionnaires have not found flavonoid intake to be inversely associated with cancer risk (85). Two prospective cohort studies in Europe found no relationship between the risk of various cancers and dietary intakes of flavones and flavonols (86, 87), catechins (88), or tea (89). In a cohort of postmenopausal women in the US, catechin intake from tea, but not fruit and vegetables, was inversely associated with the risk of rectal cancer, but not other cancers (90). Two prospective cohort studies in Finland, where average flavonoid intakes are relatively low, found that men with the highest dietary intakes of flavonols and flavones had a significantly lower risk of developing lung cancer than those with the lowest intakes (44, 45). When individual dietary flavonoids were analyzed, dietary quercetin intake, mainly from apples, was inversely associated with the risk of lung cancer; myricetin intake was inversely associated with the risk of prostate cancer (45). Tea is an important source of flavonoids (flavanols and flavonols) in some populations, but most prospective cohort studies have not found tea consumption to be inversely associated with cancer risk (reviewed in <u>91</u>). The results of <u>case-control studies</u>, which are more likely to be influenced by recall bias, are mixed. While some studies have observed lower flavonoid intakes in people diagnosed with lung (92), stomach (93, 94), and breast (95) cancer, many others have found no significant differences in flavonoid intake between cancer cases and controls (96, 97). There is limited evidence that low intakes of flavonoids from food are associated with increased risk of certain cancers, but it is not clear whether these findings are related to insufficient intakes of flavonoids or other nutrients and phytochemicals found in flavonoid-rich foods. For more information on flavonoid-rich foods and cancer, see separate articles on Fruit and Vegetables, Legumes, and Tea. Clinical trials will be necessary to determine if specific flavonoids are beneficial in the prevention or treatment of cancer; a few clinical trials are currently under way (see http://www.cancer.gov/aboutcancer/treatment/clinical-trials).

## Neurodegenerative diseases

Inflammation, oxidative stress, and transition metal accumulation appear to play a role in the pathology of several <u>neurodegenerative diseases</u>, including <u>Parkinson's disease</u> and <u>Alzheimer's</u> <u>disease (98)</u>. Because flavonoids have anti-inflammatory, antioxidant, and metal-chelating properties, scientists are interested in the neuroprotective potential of flavonoid-rich diets or individual flavonoids. At present, the extent to which various dietary flavonoids and flavonoid <u>metabolites</u> cross the blood brain barrier in humans is not known <u>(99, 100)</u>. Although flavonoid-

Flavonoids | Linus Pauling Institute | Oregon State University

rich diets and flavonoid administration have been found to prevent <u>cognitive</u> impairment associated with aging and inflammation in some animal studies (101-104), prospective cohort studies have not found consistent inverse associations between flavonoid intake and the risk of dementia or neurodegenerative disease in humans (105-109). In a cohort of Japanese-American men followed for 25-30 years, flavonoid intake from tea during midlife was not associated with the risk of Alzheimer's or other types of dementia in late life (105). Surprisingly, higher intakes of isoflavone-rich tofu during midlife were associated with cognitive impairment and brain atrophy in late life (see the article on Soy Isoflavones) (106). A prospective study of Dutch adults found that total dietary flavonoid intake was not associated with the risk of developing Parkinson's disease (107) or Alzheimer's disease (108), except in current smokers whose risk of Alzheimer's disease decreased by 50% for every 12 mg increase in daily flavonoid intake. In contrast, a study of elderly French men and women found that those with the lowest flavonoid intakes had a risk of developing dementia over the next five years that was 50% higher than those with the highest intakes (109). More recently, a study in 1,640 elderly men and women found that those with higher dietary flavonoid intake (>13.6 mg/day) had better cognitive performance at baseline and experienced significantly less age-related cognitive decline over a 10-year period than those with a lower flavonoid intake (0-10.4 mg/day) (110). Additionally, a randomized, double-blind, placebo-controlled clinical trial in 202 postmenopausal women reported that daily supplementation with 25.6 g of soy protein (containing 99 mg of isoflavones) for one year did not improve cognitive function (111). However, a randomized, double-blind, placebo-controlled, cross-over trial in 77 postmenopausal women found that six-month supplementation with 60 mg/day of isoflavones improved some measures of cognitive performance (112). Although scientists are interested in the potential of flavonoids to protect the aging brain, it is not yet clear how flavonoid consumption affects neurodegenerative disease risk in humans.

# Sources

## **Food sources**

Dietary sources of flavonoids include tea, red wine, fruit, vegetables, and legumes. Individual flavonoid intakes may vary considerably depending on whether tea, red wine, soy products, or fruit and vegetables are commonly consumed (reviewed in <u>3</u>). Although individual flavonoid intakes may vary, total flavonoid intakes in Western populations appear to average about 150-200 mg/day (<u>3</u>, <u>113</u>). Information on the flavonoid content of some flavonoid-rich foods is presented in <u>Table 2</u> and <u>Table 3</u>. These values should be considered approximate since a number of factors may affect the flavonoid content of foods, including agricultural practices, environmental factors, ripening, processing, storing, and cooking. For more information about the flavonoid content of selected foods. For information on the isoflavone content of soy foods, see the separate article on <u>Soy Isoflavones</u> or the USDA database for the <u>isoflavone</u> content of selected foods.

Table 2. Anthocyanin, Flavanol, and Proanthocyanidin Content of Selected Foods (mg/100 g or 100 mL*) (3, 129-135)					
Anthocyanin-rich Foods Anthocyanins Flavanols Proanthocyanidi					
Blackberry	89-211	13-19	6-47		
Blueberry	67-183	1	88-261		
Grapes, red	25-92	2	44-76		
Raspberries (red)	10-84	9	5-59		
Strawberry	15-75	-	97-183		
Red wine	1-35	1-55	24-70		
Plum	2-25	1-6	106-334		

http://lpi.oregonstate.edu/mic/dietary-factors/phytochemicals/flavonoids[2016/01/28 21:36:31]

Flavonoids | Linus Pauling Institute | Oregon State University

Red cabbage	25	0	-
Red onion	13- 25	-	-
Blood orange juice	3-10	-	-
Flavanol-rich Foods	Anthocyanins	Flavanols	Proanthocyaniding
Green tea	-	24-216	-
Black tea	-	5-158	4
Chocolate, dark	-	43-63	90-322
Apple, red delicious with peel	1-4	2-12	89-148
Apricot	-	10-25	8-13
Flavone-rich Foods	Anthocyanins	Flavanols	Proanthocyaniding
Parsley, fresh	-	-	-
Thyme, fresh	-	-	-
Celery hearts, green	-	-	-
Celery	-	-	-
Oregano, fresh	-	-	-
Chili peppers, green	-	-	-
Flavanone-rich Foods	Anthocyanins	Flavanols	Proanthocyaniding
Lemon juice, fresh	-	-	-
Grapefruit juice, fresh	-	-	-
Orange juice, fresh	-	-	-
Grapefruit, fresh	-	-	-
Orange, fresh	-	-	-
Flavonol-rich Foods	Anthocyanins	Flavanols	Proanthocyaniding
Onion, yellow	-	0	-
Kale	-	-	-
Leek	-	0	-
Broccoli		0	

\*per 100 g (fresh weight) or 100 mL (liquids); 100 grams is equivalent to about 3.5 ounces; 100 mL is equivalent to about 3.5 fluid ounces.

Table 3. Flavone, Flavonol, and Flavanone Content of Selected Foods (mg/100 g or 100 mL*) <u>(3, 129-135)</u>			
Anthocyanin-rich Foods	Flavones	Flavonols	Flavanones
Blackberry	-	0-2	-
Blueberry	-	2-16	-
Grapes, red	-	3-4	-
Blackberry Blueberry	-	0-2 2-16	-

 $http://lpi.oregonstate.edu/mic/dietary-factors/phytochemicals/flavonoids [2016/01/28\ 21:36:31]$ 

Flavonoids | Linus Pauling Institute | Oregon State University

Raspberries (red)	-	1	-
Strawberry	-	1-4	-
Red wine	0	2-30	-
Plum	0	1-2	-
Red cabbage	0-1	0-1	-
Red onion	0	4-100	-
Blood orange juice	-	-	10-22
Flavanol-rich Foods	Flavones	Flavonols	Flavanones
Green tea	0-1	3-9	-
Black tea	0	1-7	-
Chocolate, dark	-	-	-
Apple, red delicious with peel	0	2-6	-
Apricot	0	2-5	-
Flavone-rich Foods	Flavones	Flavonols	Flavanones
Parsley, fresh	24-634	8-10	-
Thyme, fresh	56	0	-
Celery hearts, green	23	-	-
Celery	0-15	4	-
Oregano, fresh	2-7	0	-
Chili peppers, green	5	13-21	-
Flavanone-rich Foods	Flavones	Flavonols	Flavanones
Lemon juice, fresh	0	0-2	2-175
Grapefruit juice, fresh	0	0	10-104
Orange juice, fresh	0-1	0	5-47
Grapefruit, fresh	-	1	55
Orange, fresh	-	-	42-53
Flavonol-rich Foods	Flavones	Flavonols	Flavanones
Onion, yellow	0	3-120	-
Kale	0	30-60	-
Leek	0	3-22	-
Broccoli	0	4-13	_

\*per 100 g (fresh weight) or 100 mL (liquids); 100 grams is equivalent to about 3.5 ounces; 100 mL is equivalent to about 3.5 fluid ounces.

## **Supplements**

### Anthocyanins

Bilberry, elderberry, black currant, blueberry, red grape, and mixed berry extracts that are rich in

anthocyanins are available as dietary supplements without a prescription in the US. The anthocyanin content of these products may vary considerably. Standardized extracts that list the amount of anthocyanins per dose are available.

#### Flavanols

Numerous tea extracts are available in the US as dietary supplements and may be labeled as tea catechins or tea polyphenols. Green tea extracts are the most commonly marketed, but black and oolong tea extracts are also available. Green tea extracts generally have higher levels of catechins (flavanol monomers), while black tea extracts are richer in theaflavins and thearubigins (flavanol polymers found in tea). Oolong tea extracts fall somewhere in between green and black tea extracts with respect to their flavanol content. Some tea extracts contain caffeine, while others are decaffeinated. Flavanol and caffeine content vary considerably among different products, so it is important to check the label or consult the manufacturer to determine the amounts of flavanols and caffeine that would be consumed daily with each supplement. For more information on tea flavanols, see the article on Tea.

#### Flavanones

Citrus bioflavonoid supplements may contain glycosides of hesperetin (hesperidin), naringenin (naringin), and eriodictyol (eriocitrin). Hesperidin is also available in hesperidin-complex supplements (114).

#### Flavones

The peels of citrus fruit are rich in polymethoxylated flavones: tangeretin, nobiletin, and sinensetin (3). Although dietary intakes of these naturally occurring flavones are generally low, they are often present in citrus bioflavonoid supplements.

#### Flavonols

The flavonol aglycone, quercetin, and its glycoside rutin are available as dietary supplements without a prescription in the US. Other names for rutin include rutinoside, quercetin-3-rutinoside, and sophorin (114). Citrus bioflavonoid supplements may also contain quercetin or rutin.

### Safety

### **Adverse effects**

No adverse effects have been associated with high dietary intakes of flavonoids from plant-based foods. This lack of adverse effects may be explained by the relatively low bioavailability and rapid metabolism and elimination of most flavonoids.

#### Quercetin

Some men taking quercetin supplements (1,000 mg/day for one month) reported nausea, headache, or tingling of the extremities (115). Some cancer patients given intravenous quercetin in a phase I clinical trial reported nausea, vomiting, sweating, flushing, and dyspnea (difficulty breathing) (116). Intravenous administration of quercetin at doses of 945 mg/m<sup>2</sup> or more was associated with renal (kidney) toxicity in that trial.

Tea extracts

There have been several reports of hepatotoxicity (liver toxicity) following consumption of supplements containing tea (Camellia sinensis) extracts (<u>117, 118)</u>. In clinical trials of caffeinated

green tea extracts, cancer patients who took 6 g/day in 3-6 divided doses have reported mild to moderate gastrointestinal side effects, including nausea, vomiting, abdominal pain, and diarrhea (119, 120). Central nervous system symptoms, including agitation, restlessness, insomnia, tremors, dizziness, and confusion, have also been reported. In one case, confusion was severe enough to require hospitalization (119). These side effects were likely related to the caffeine in the green tea extract (120). In a four-week clinical trial that assessed the safety of decaffeinated green tea extracts (800 mg/day of EGCG) in healthy individuals, a few of the participants reported mild nausea, stomach upset, dizziness, or muscle pain (121).

## **Pregnancy and lactation**

The safety of flavonoid supplements in pregnancy and lactation has not been established (114).

## **Drug interactions**

### Inhibition of CYP 3A4 by grapefruit juice and flavonoids

As little as 200 mL (7 fluid ounces) of grapefruit juice has been found to irreversibly inhibit the intestinal drug metabolizing <u>enzyme</u>, cytochrome P450 (CYP) 3A4 (122). Although the most potent inhibitors of CYP3A4 in grapefruit are thought to be furanocoumarins, particularly dihydroxybergamottin, the flavonoids naringenin and guercetin have also been found to inhibit CYP3A4 *in vitro*. Inhibition of intestinal CYP3A4 can increase the bioavailability and the risk of toxicity of a number of drugs, including but not limited to HMG-CoA reductase inhibitors (atorvastatin, lovastatin, and simvastatin), calcium channel antagonists (felodipine, nicardipine, nisoldipine, nitrendipine, and verapamil), anti-arrhythmic agents (amiodarone), HIV protease inhibitors (saquinavir), immunosuppressants (cyclosporine), antihistamines (terfenadine), gastrointestinal stimulants (cisapride), benzodiazepines (diazepam, midazolam, and triazolam), anticonvulsants (carbamazepine), anxiolytics (buspirone) serotonin specific reuptake inhibitors (sertraline), and drugs used to treat erectile dysfunction (sildenafil) (123). Grapefruit juice may reduce the therapeutic effect of the angiotensin II receptor antagonist, losartan. Because of the potential for adverse drug interactions, some clinicians recommend that people taking medications that undergo extensive presystemic metabolism by CYP3A4 avoid consuming grapefruit juice altogether to avoid potential toxicities (122).

### Inhibition of P-glycoprotein by grapefruit juice and flavonoids

P-glycoprotein is an efflux transporter that decreases the absorption of a number of drugs. There is some evidence that the consumption of grapefruit juice inhibits the activity of P-glycoprotein (122). Quercetin, naringenin, and the green tea flavanol, epigallocatechin gallate (EGCG), have been found to inhibit the efflux activity of P-glycoprotein in cultured cells (124). Thus, very high or supplemental intakes of these flavonoids could potentially increase flavonoid bioavailability, potentially increasing the toxicity of drugs that are substrates of P-glycoprotein. Drugs known to be substrates of P-glycoprotein include digoxin, antihypertensive agents, antiarrhythmic agents, chemotherapeutic (anticancer) agents, antifungal agents, HIV protease inhibitors, immunosuppressive agents, H2 receptor antagonists, some antibiotics, and others (reviewed in 125).

### Anticoagulant and antiplatelet drugs

High intakes of flavonoids from purple grape juice (500 mL/day) and dark chocolate (235 mg/day of flavanols) have been found to inhibit <u>platelet</u> aggregation in *ex vivo* assays (69-71, 75). Theoretically, high intakes of flavonoids (e.g., from supplements) could increase the risk of bleeding when taken with anticoagulant drugs, such as warfarin (Coumadin), and antiplatelet drugs, such as clopidogrel (Plavix), dipyridamole (Persantine), non-steroidal anti-inflamatory drugs (NSAIDs), aspirin, and others.

# **Nutrient interactions**

#### Nonheme iron

Flavonoids can bind nonheme iron, inhibiting its intestinal absorption. Nonheme iron is the principal form of iron in plant foods, dairy products, and iron supplements. The consumption of one cup of tea or cocoa with a meal has been found to decrease the absorption of nonheme iron in that meal by about 70% (126, 127). To maximize iron absorption from a meal or iron supplements, flavonoid-rich beverages or flavonoid supplements should not be taken at the same time.

Vitamin C

Studies in cell culture indicate that a number of flavonoids inhibit the transport of <u>vitamin C</u> into cells, and supplementation of rats with quercetin and vitamin C decreased the intestinal absorption of vitamin C (128). More research is needed to determine the significance of these findings in humans.

### **Authors and Reviewers**

Originally written in 2005 by: Jane Higdon, Ph.D. Linus Pauling Institute Oregon State University

Updated in June 2008 by: Victoria J. Drake, Ph.D. Linus Pauling Institute Oregon State University

Reviewed in June 2008 by: Roderick H. Dashwood, Ph.D. Director, Cancer Chemoprotection Program, Linus Pauling Institute Professor of Environmental & Molecular Toxicology Leader, Environmental Mutagenesis & Carcinogenesis Core, Environmental Health Sciences Center

Oregon State University

Copyright 2005-2016 Linus Pauling Institute

### References

1. Beecher GR. Overview of dietary flavonoids: nomenclature, occurrence and intake. J Nutr. 2003;133(10):3248S-3254S. (<u>PubMed)</u>

2. Williamson G. Common features in the pathways of absorption and metabolism of flavonoids. In: Meskin MS, R. BW, Davies AJ, Lewis DS, Randolph RK, eds. Phytochemicals: Mechanisms of Action. Boca Raton: CRC Press; 2004:21-33.

3. Manach C, Scalbert A, Morand C, Remesy C, Jimenez L. Polyphenols: food sources and bioavailability. Am J Clin Nutr. 2004;79(5):727-747. (PubMed)

4. Setchell KD, Brown NM, Lydeking-Olsen E. The clinical importance of the metabolite equol-a clue to the effectiveness of soy and its isoflavones. J Nutr. 2002;132(12):3577-3584. (PubMed)

5. Yuan JP, Wang JH, Liu X. Metabolism of dietary soy isoflavones to equol by human intestinal microflora--implications for health. Mol Nutr Food Res. 2007;51(7):765-781. (PubMed)

6. Manach C, Williamson G, Morand C, Scalbert A, Remesy C. Bioavailability and bioefficacy of polyphenols in humans. I. Review of 97 bioavailability studies. Am J Clin Nutr. 2005;81(1 Suppl):230S-242S. (PubMed)

7. Williams RJ, Spencer JP, Rice-Evans C. Flavonoids: antioxidants or signalling molecules? Free Radic Biol Med. 2004;36(7):838-849. (PubMed)

8. Kroon PA, Clifford MN, Crozier A, et al. How should we assess the effects of exposure to dietary polyphenols in vitro? Am J Clin Nutr. 2004;80(1):15-21. (PubMed)

9. Heijnen CG, Haenen GR, van Acker FA, van der Vijgh WJ, Bast A. Flavonoids as peroxynitrite scavengers: the role of the hydroxyl groups. Toxicol In Vitro. 2001;15(1):3-6. (PubMed)

10. Chun OK, Kim DO, Lee CY. Superoxide radical scavenging activity of the major polyphenols in fresh plums. J Agric Food Chem. 2003;51(27):8067-8072. (PubMed)

11. Frei B, Higdon JV. Antioxidant activity of tea polyphenols in vivo: evidence from animal studies. J Nutr. 2003;133(10):3275S-3284S. (<u>PubMed)</u>

12. Lotito SB, Frei B. Consumption of flavonoid-rich foods and increased plasma antioxidant capacity in humans: cause, consequence, or epiphenomenon? Free Radic Biol Med. 2006;41(12):1727-1746. (PubMed)

13. Mira L, Fernandez MT, Santos M, Rocha R, Florencio MH, Jennings KR. Interactions of flavonoids with iron and copper ions: a mechanism for their antioxidant activity. Free Radic Res. 2002;36(11):1199-1208. (PubMed)

14. Cheng IF, Breen K. On the ability of four flavonoids, baicilein, luteolin, naringenin, and quercetin, to suppress the Fenton reaction of the iron-ATP complex. Biometals. 2000;13(1):77-83. (PubMed)

15. Spencer JP, Rice-Evans C, Williams RJ. Modulation of pro-survival Akt/protein kinase B and ERK1/2 signaling cascades by quercetin and its in vivo metabolites underlie their action on neuronal viability. J Biol Chem. 2003;278(37):34783-34793. (PubMed)

16. Spencer JP, Schroeter H, Crossthwaithe AJ, Kuhnle G, Williams RJ, Rice-Evans C. Contrasting influences of glucuronidation and O-methylation of epicatechin on hydrogen peroxide-induced cell death in neurons and fibroblasts. Free Radic Biol Med. 2001;31(9):1139-1146. (PubMed)

17. Hou Z, Lambert JD, Chin KV, Yang CS. Effects of tea polyphenols on signal transduction pathways related to cancer chemoprevention. Mutat Res. 2004;555(1-2):3-19. (PubMed)

18. Lambert JD, Yang CS. Mechanisms of cancer prevention by tea constituents. J Nutr. 2003;133(10):3262S-3267S. (PubMed)

19. Kong AN, Owuor E, Yu R, et al. Induction of xenobiotic enzymes by the MAP kinase pathway and the antioxidant or electrophile response element (ARE/EpRE). Drug Metab Rev. 2001;33(3-4):255-271. (PubMed)

20. Walle UK, Walle T. Induction of human UDP-glucuronosyltransferase UGT1A1 by flavonoidsstructural requirements. Drug Metab Dispos. 2002;30(5):564-569. (<u>PubMed</u>)

21. Chen JJ, Ye ZQ, Koo MW. Growth inhibition and cell cycle arrest effects of epigallocatechin gallate in the NBT-II bladder tumour cell line. BJU Int. 2004;93(7):1082-1086. (<u>PubMed</u>)

22. Wang W, VanAlstyne PC, Irons KA, Chen S, Stewart JW, Birt DF. Individual and interactive effects of apigenin analogs on G2/M cell-cycle arrest in human colon carcinoma cell lines. Nutr Cancer. 2004;48(1):106-114. (PubMed)

23. Stewart ZA, Westfall MD, Pietenpol JA. Cell-cycle dysregulation and anticancer therapy. Trends Pharmacol Sci. 2003;24(3):139-145. (PubMed)

24. Sah JF, Balasubramanian S, Eckert RL, Rorke EA. Epigallocatechin-3-gallate inhibits epidermal growth factor receptor signaling pathway. Evidence for direct inhibition of ERK1/2 and AKT kinases. J Biol Chem. 2004;279(13):12755-12762. (PubMed)

25. Kavanagh KT, Hafer LJ, Kim DW, et al. Green tea extracts decrease carcinogen-induced mammary tumor burden in rats and rate of breast cancer cell proliferation in culture. J Cell Biochem. 2001;82(3):387-398. (PubMed)

26. Ramos S. Effects of dietary flavonoids on apoptotic pathways related to cancer chemoprevention. J Nutr Biochem. 2007;18(7):427-442. (PubMed)

27. Bagli E, Stefaniotou M, Morbidelli L, et al. Luteolin inhibits vascular endothelial growth factorinduced angiogenesis; inhibition of endothelial cell survival and proliferation by targeting phosphatidylinositol 3'-kinase activity. Cancer Res. 2004;64(21):7936-7946. (PubMed)

28. Kim MH. Flavonoids inhibit VEGF/bFGF-induced angiogenesis in vitro by inhibiting the matrix-degrading proteases. J Cell Biochem. 2003;89(3):529-538. (PubMed)

29. O'Leary KA, de Pascual-Tereasa S, Needs PW, Bao YP, O'Brien NM, Williamson G. Effect of flavonoids and vitamin E on cyclooxygenase-2 (COX-2) transcription. Mutat Res. 2004;551(1-2):245-254. (PubMed)

30. Sakata K, Hirose Y, Qiao Z, Tanaka T, Mori H. Inhibition of inducible isoforms of cyclooxygenase and nitric oxide synthase by flavonoid hesperidin in mouse macrophage cell line. Cancer Lett. 2003;199(2):139-145. (PubMed)

31. Cho SY, Park SJ, Kwon MJ, et al. Quercetin suppresses proinflammatory cytokines production through MAP kinases and NF-kappaB pathway in lipopolysaccharide-stimulated macrophage. Mol Cell Biochem. 2003;243(1-2):153-160. (PubMed)

32. Steele VE, Hawk ET, Viner JL, Lubet RA. Mechanisms and applications of non-steroidal antiinflammatory drugs in the chemoprevention of cancer. Mutat Res. 2003;523-524:137-144. (PubMed)

33. Blake GJ, Ridker PM. C-reactive protein and other inflammatory risk markers in acute coronary syndromes. J Am Coll Cardiol. 2003;41(4 Suppl S):37S-42S. (PubMed)

34. Choi JS, Choi YJ, Park SH, Kang JS, Kang YH. Flavones mitigate tumor necrosis factor-alphainduced adhesion molecule upregulation in cultured human endothelial cells: role of nuclear factor-kappa B. J Nutr. 2004;134(5):1013-1019. (<u>PubMed</u>)

35. Ludwig A, Lorenz M, Grimbo N, et al. The tea flavonoid epigallocatechin-3-gallate reduces cytokine-induced VCAM-1 expression and monocyte adhesion to endothelial cells. Biochem Biophys Res Commun. 2004;316(3):659-665. (PubMed)

36. Stocker R, Keaney JF, Jr. Role of oxidative modifications in atherosclerosis. Physiol Rev. 2004;84(4):1381-1478. (PubMed)

37. Anter E, Thomas SR, Schulz E, Shapira OM, Vita JA, Keaney JF, Jr. Activation of endothelial nitric-oxide synthase by the p38 MAPK in response to black tea polyphenols. J Biol Chem. 2004;279(45):46637-46643. (PubMed)

38. Duffy SJ, Vita JA. Effects of phenolics on vascular endothelial function. Curr Opin Lipidol. 2003;14(1):21-27. (PubMed)

39. Deana R, Turetta L, Donella-Deana A, et al. Green tea epigallocatechin-3-gallate inhibits platelet signalling pathways triggered by both proteolytic and non-proteolytic agonists. Thromb Haemost. 2003;89(5):866-874. (PubMed)

40. Bucki R, Pastore JJ, Giraud F, Sulpice JC, Janmey PA. Flavonoid inhibition of platelet procoagulant activity and phosphoinositide synthesis. J Thromb Haemost. 2003;1(8):1820-1828. (<u>PubMed</u>)

41. Hubbard GP, Wolffram S, Lovegrove JA, Gibbins JM. The role of polyphenolic compounds in the diet as inhibitors of platelet function. Proc Nutr Soc. 2003;62(2):469-478. (PubMed)

42. Geleijnse JM, Launer LJ, Van der Kuip DA, Hofman A, Witteman JC. Inverse association of tea and flavonoid intakes with incident myocardial infarction: the Rotterdam Study. Am J Clin Nutr. 2002;75(5):880-886. (PubMed)

43. Hertog MG, Feskens EJ, Kromhout D. Antioxidant flavonols and coronary heart disease risk. Lancet. 1997;349(9053):699. (PubMed)

44. Hirvonen T, Pietinen P, Virtanen M, et al. Intake of flavonols and flavones and risk of coronary heart disease in male smokers. Epidemiology. 2001;12(1):62-67. (PubMed)

45. Knekt P, Kumpulainen J, Jarvinen R, et al. Flavonoid intake and risk of chronic diseases. Am J Clin Nutr. 2002;76(3):560-568. (PubMed)

46. Yochum L, Kushi LH, Meyer K, Folsom AR. Dietary flavonoid intake and risk of cardiovascular disease in postmenopausal women. Am J Epidemiol. 1999;149(10):943-949. (PubMed)

47. Hertog MG, Sweetnam PM, Fehily AM, Elwood PC, Kromhout D. Antioxidant flavonols and ischemic heart disease in a Welsh population of men: the Caerphilly Study. Am J Clin Nutr. 1997;65(5):1489-1494. (PubMed)

48. Rimm EB, Katan MB, Ascherio A, Stampfer MJ, Willett WC. Relation between intake of flavonoids and risk for coronary heart disease in male health professionals. Ann Intern Med. 1996;125(5):384-389. (PubMed)

49. Sesso HD, Gaziano JM, Liu S, Buring JE. Flavonoid intake and the risk of cardiovascular disease in women. Am J Clin Nutr. 2003;77(6):1400-1408. (PubMed)

50. Mink PJ, Scrafford CG, Barraj LM, et al. Flavonoid intake and cardiovascular disease mortality: a prospective study in postmenopausal women. Am J Clin Nutr. 2007;85(3):895-909. (PubMed)

51. Lin J, Rexrode KM, Hu F, et al. Dietary intakes of flavonols and flavones and coronary heart disease in US women. Am J Epidemiol. 2007;165(11):1305-1313. (PubMed)

52. Keli SO, Hertog MG, Feskens EJ, Kromhout D. Dietary flavonoids, antioxidant vitamins, and incidence of stroke: the Zutphen study. Arch Intern Med. 1996;156(6):637-642. (PubMed)

53. Hirvonen T, Virtamo J, Korhonen P, Albanes D, Pietinen P. Intake of flavonoids, carotenoids, vitamins C and E, and risk of stroke in male smokers. Stroke. 2000;31(10):2301-2306. (PubMed)

54. Knekt P, Isotupa S, Rissanen H, et al. Quercetin intake and the incidence of cerebrovascular disease. Eur J Clin Nutr. 2000;54(5):415-417. (PubMed)

55. Liu RH. Health benefits of fruit and vegetables are from additive and synergistic combinations of phytochemicals. Am J Clin Nutr. 2003;78(3 Suppl):517S-520S. (PubMed)

56. Vita JA. Tea consumption and cardiovascular disease: effects on endothelial function. J Nutr. 2003;133(10):3293S-3297S. (PubMed)

57. Duffy SJ, Keaney JF, Jr., Holbrook M, et al. Short- and long-term black tea consumption reverses endothelial dysfunction in patients with coronary artery disease. Circulation. 2001;104(2):151-156. (PubMed)

58. Hodgson JM, Puddey IB, Burke V, Watts GF, Beilin LJ. Regular ingestion of black tea improves brachial artery vasodilator function. Clin Sci (Lond). 2002;102(2):195-201. (PubMed)

59. Stein JH, Keevil JG, Wiebe DA, Aeschlimann S, Folts JD. Purple grape juice improves endothelial function and reduces the susceptibility of LDL cholesterol to oxidation in patients with coronary artery disease. Circulation. 1999;100(10):1050-1055. (PubMed)

60. Engler MB, Engler MM, Chen CY, et al. Flavonoid-rich dark chocolate improves endothelial function and increases plasma epicatechin concentrations in healthy adults. J Am Coll Nutr. 2004;23(3):197-204. (PubMed)

61. Wang-Polagruto JF, Villablanca AC, Polagruto JA, et al. Chronic consumption of flavanol-rich cocoa improves endothelial function and decreases vascular cell adhesion molecule in hypercholesterolemic postmenopausal women. J Cardiovasc Pharmacol. 2006;47 Suppl 2:S177-186; discussion S206-179. (PubMed)

62. Balzer J, Rassaf T, Heiss C, et al. Sustained benefits in vascular function through flavanolcontaining cocoa in medicated diabetic patients a double-masked, randomized, controlled trial. J Am Coll Cardiol. 2008;51(22):2141-2149. (PubMed)

63. Schroeter H, Heiss C, Balzer J, et al. (-)-Epicatechin mediates beneficial effects of flavanolrich cocoa on vascular function in humans. Proc Natl Acad Sci U S A. 2006;103(4):1024-1029. (PubMed)

64. Taubert D, Roesen R, Lehmann C, Jung N, Schomig E. Effects of low habitual cocoa intake on blood pressure and bioactive nitric oxide: a randomized controlled trial. JAMA. 2007;298(1):49-60. (PubMed)

65. Freese R, Vaarala O, Turpeinen AM, Mutanen M. No difference in platelet activation or inflammation markers after diets rich or poor in vegetables, berries and apple in healthy subjects. Eur J Nutr. 2004;43(3):175-182. (PubMed)

66. Janssen K, Mensink RP, Cox FJ, et al. Effects of the flavonoids quercetin and apigenin on hemostasis in healthy volunteers: results from an in vitro and a dietary supplement study. Am J Clin Nutr. 1998;67(2):255-262. (PubMed)

67. Duffy SJ, Vita JA, Holbrook M, Swerdloff PL, Keaney JF, Jr. Effect of acute and chronic tea consumption on platelet aggregation in patients with coronary artery disease. Arterioscler Thromb Vasc Biol. 2001;21(6):1084-1089. (PubMed)

68. Hodgson JM, Puddey IB, Burke V, Beilin LJ, Mori TA, Chan SY. Acute effects of ingestion of black tea on postprandial platelet aggregation in human subjects. Br J Nutr. 2002;87(2):141-145. (PubMed)

69. Freedman JE, Parker C, 3rd, Li L, et al. Select flavonoids and whole juice from purple grapes inhibit platelet function and enhance nitric oxide release. Circulation. 2001;103(23):2792-2798. (PubMed)

70. Keevil JG, Osman HE, Reed JD, Folts JD. Grape juice, but not orange juice or grapefruit juice, inhibits human platelet aggregation. J Nutr. 2000;130(1):53-56. (PubMed)

71. Polagruto JA, Schramm DD, Wang-Polagruto JF, Lee L, Keen CL. Effects of flavonoid-rich beverages on prostacyclin synthesis in humans and human aortic endothelial cells: association with ex vivo platelet function. J Med Food. 2003;6(4):301-308. (PubMed)

72. Innes AJ, Kennedy G, McLaren M, Bancroft AJ, Belch JJ. Dark chocolate inhibits platelet aggregation in healthy volunteers. Platelets. 2003;14(5):325-327. (PubMed)

73. Rein D, Paglieroni TG, Pearson DA, et al. Cocoa and wine polyphenols modulate platelet activation and function. J Nutr. 2000;130(8S Suppl):2120S-2126S. (PubMed)

74. Rein D, Paglieroni TG, Wun T, et al. Cocoa inhibits platelet activation and function. Am J Clin Nutr. 2000;72(1):30-35. (PubMed)

75. Murphy KJ, Chronopoulos AK, Singh I, et al. Dietary flavanols and procyanidin oligomers from cocoa (Theobroma cacao) inhibit platelet function. Am J Clin Nutr. 2003;77(6):1466-1473. (PubMed)

76. Yang CS, Yang GY, Landau JM, Kim S, Liao J. Tea and tea polyphenols inhibit cell hyperproliferation, lung tumorigenesis, and tumor progression. Exp Lung Res. 1998;24(4):629-639. (PubMed)

77. Balasubramanian S, Govindasamy S. Inhibitory effect of dietary flavonol quercetin on 7,12dimethylbenz[a]anthracene-induced hamster buccal pouch carcinogenesis. Carcinogenesis. 1996;17(4):877-879. (PubMed)

78. Li ZG, Shimada Y, Sato F, et al. Inhibitory effects of epigallocatechin-3-gallate on Nnitrosomethylbenzylamine-induced esophageal tumorigenesis in F344 rats. Int J Oncol. 2002;21(6):1275-1283. (PubMed)

79. Yamane T, Nakatani H, Kikuoka N, et al. Inhibitory effects and toxicity of green tea polyphenols for gastrointestinal carcinogenesis. Cancer. 1996;77(8 Suppl):1662-1667. (PubMed)

80. Guo JY, Li X, Browning JD, Jr., et al. Dietary soy isoflavones and estrone protect ovariectomized ERalphaKO and wild-type mice from carcinogen-induced colon cancer. J Nutr. 2004;134(1):179-182. (PubMed)

81. Huang MT, Xie JG, Wang ZY, et al. Effects of tea, decaffeinated tea, and caffeine on UVB light-induced complete carcinogenesis in SKH-1 mice: demonstration of caffeine as a biologically important constituent of tea. Cancer Res. 1997;57(13):2623-2629. (PubMed)

82. Gupta S, Hastak K, Ahmad N, Lewin JS, Mukhtar H. Inhibition of prostate carcinogenesis in TRAMP mice by oral infusion of green tea polyphenols. Proc Natl Acad Sci U S A. 2001;98(18):10350-10355. (PubMed)

83. Haddad AQ, Venkateswaran V, Viswanathan L, Teahan SJ, Fleshner NE, Klotz LH. Novel antiproliferative flavonoids induce cell cycle arrest in human prostate cancer cell lines. Prostate Cancer Prostatic Dis. 2006;9(1):68-76. (PubMed)

84. Yamagishi M, Natsume M, Osakabe N, et al. Effects of cacao liquor proanthocyanidins on PhIP-induced mutagenesis in vitro, and in vivo mammary and pancreatic tumorigenesis in female Sprague-Dawley rats. Cancer Lett. 2002;185(2):123-130. (PubMed)

85. Ross JA, Kasum CM. Dietary flavonoids: bioavailability, metabolic effects, and safety. Annu Rev Nutr. 2002;22:19-34. (PubMed)

86. Goldbohm RA, Van den Brandt PA, Hertog MG, Brants HA, Van Poppel G. Flavonoid intake and risk of cancer: a prospective cohort study. Am J Epidemiol. 1995;41:s61.

87. Hertog MG, Feskens EJ, Hollman PC, Katan MB, Kromhout D. Dietary flavonoids and cancer risk in the Zutphen Elderly Study. Nutr Cancer. 1994;22(2):175-184. (PubMed)

88. Arts IC, Hollman PC, Bueno De Mesquita HB, Feskens EJ, Kromhout D. Dietary catechins and epithelial cancer incidence: the Zutphen elderly study. Int J Cancer. 2001;92(2):298-302. (PubMed)

89. Goldbohm RA, Hertog MG, Brants HA, van Poppel G, van den Brandt PA. Consumption of black tea and cancer risk: a prospective cohort study. J Natl Cancer Inst. 1996;88(2):93-100. (<u>PubMed)</u>

90. Arts IC, Jacobs DR, Jr., Gross M, Harnack LJ, Folsom AR. Dietary catechins and cancer incidence among postmenopausal women: the Iowa Women's Health Study (United States). Cancer Causes Control. 2002;13(4):373-382. (PubMed)

91. Higdon JV, Frei B. Tea catechins and polyphenols: health effects, metabolism, and antioxidant functions. Crit Rev Food Sci Nutr. 2003;43(1):89-143. (PubMed)

92. De Stefani E, Ronco A, Mendilaharsu M, Deneo-Pellegrini H. Diet and risk of cancer of the upper aerodigestive tract--II. Nutrients. Oral Oncol. 1999;35(1):22-26. (PubMed)

93. Garcia-Closas R, Gonzalez CA, Agudo A, Riboli E. Intake of specific carotenoids and flavonoids and the risk of gastric cancer in Spain. Cancer Causes Control. 1999;10(1):71-75. (PubMed)

94. Lagiou P, Samoli E, Lagiou A, et al. Flavonoids, vitamin C and adenocarcinoma of the stomach. Cancer Causes Control. 2004;15(1):67-72. (<u>PubMed)</u>

95. Peterson J, Lagiou P, Samoli E, et al. Flavonoid intake and breast cancer risk: a case--control study in Greece. Br J Cancer. 2003;89(7):1255-1259. (PubMed)

96. Garcia R, Gonzalez CA, Agudo A, Riboli E. High intake of specific carotenoids and flavonoids does not reduce the risk of bladder cancer. Nutr Cancer. 1999;35(2):212-214. (PubMed)

97. Garcia-Closas R, Agudo A, Gonzalez CA, Riboli E. Intake of specific carotenoids and flavonoids and the risk of lung cancer in women in Barcelona, Spain. Nutr Cancer. 1998;32(3):154-158. (PubMed)

98. Ramassamy C. Emerging role of polyphenolic compounds in the treatment of neurodegenerative diseases: a review of their intracellular targets. Eur J Pharmacol. 2006;545(1):51-64. (PubMed)

99. Youdim KA, Qaiser MZ, Begley DJ, Rice-Evans CA, Abbott NJ. Flavonoid permeability across an in situ model of the blood-brain barrier. Free Radic Biol Med. 2004;36(5):592-604. (PubMed)

100. Schmitt-Schillig S, Schaffer S, Weber CC, Eckert GP, Muller WE. Flavonoids and the aging brain. J Physiol Pharmacol. 2005;56 Suppl 1:23-36. (PubMed)

101. Goyarzu P, Malin DH, Lau FC, et al. Blueberry supplemented diet: effects on object recognition memory and nuclear factor-kappa B levels in aged rats. Nutr Neurosci. 2004;7(2):75-83. (PubMed)

102. Joseph JA, Denisova NA, Arendash G, et al. Blueberry supplementation enhances signaling and prevents behavioral deficits in an Alzheimer disease model. Nutr Neurosci. 2003;6(3):153-162. (PubMed)

103. Joseph JA, Shukitt-Hale B, Denisova NA, et al. Reversals of age-related declines in neuronal signal transduction, cognitive, and motor behavioral deficits with blueberry, spinach, or strawberry dietary supplementation. J Neurosci. 1999;19(18):8114-8121. (PubMed)

104. Patil CS, Singh VP, Satyanarayan PS, Jain NK, Singh A, Kulkarni SK. Protective effect of flavonoids against aging- and lipopolysaccharide-induced cognitive impairment in mice. Pharmacology. 2003;69(2):59-67. (PubMed)

105. Laurin D, Masaki KH, Foley DJ, White LR, Launer LJ. Midlife dietary intake of antioxidants and risk of late-life incident dementia: the Honolulu-Asia Aging Study. Am J Epidemiol. 2004;159(10):959-967. (PubMed)

106. White LR, Petrovitch H, Ross GW, et al. Brain aging and midlife tofu consumption. J Am Coll Nutr. 2000;19(2):242-255. (PubMed)

107. de Rijk MC, Breteler MM, den Breeijen JH, et al. Dietary antioxidants and Parkinson disease. The Rotterdam Study. Arch Neurol. 1997;54(6):762-765. (PubMed)

108. Engelhart MJ, Geerlings MI, Ruitenberg A, et al. Dietary intake of antioxidants and risk of Alzheimer disease. JAMA. 2002;287(24):3223-3229. (PubMed)

109. Commenges D, Scotet V, Renaud S, Jacqmin-Gadda H, Barberger-Gateau P, Dartigues JF. Intake of flavonoids and risk of dementia. Eur J Epidemiol. 2000;16(4):357-363. (<u>PubMed</u>)

110. Letenneur L, Proust-Lima C, Le Gouge A, Dartigues JF, Barberger-Gateau P. Flavonoid intake and cognitive decline over a 10-year period. Am J Epidemiol. 2007;165(12):1364-1371. (PubMed)

111. Kreijkamp-Kaspers S, Kok L, Grobbee DE, et al. Effect of soy protein containing isoflavones on cognitive function, bone mineral density, and plasma lipids in postmenopausal women: a randomized controlled trial. JAMA. 2004;292(1):65-74. (PubMed)

112. Casini ML, Marelli G, Papaleo E, Ferrari A, D'Ambrosio F, Unfer V. Psychological assessment of the effects of treatment with phytoestrogens on postmenopausal women: a randomized, double-blind, crossover, placebo-controlled study. Fertil Steril. 2006;85(4):972-978. (PubMed)

113. Gu L, Kelm MA, Hammerstone JF, et al. Concentrations of proanthocyanidins in common foods and estimations of normal consumption. J Nutr. 2004;134(3):613-617. (PubMed)

114. Hendler SS, Rorvik DR, eds. PDR for Nutritional Supplements. Montvale: Medical Economics Company, Inc; 2001.

115. Shoskes DA, Zeitlin SI, Shahed A, Rajfer J. Quercetin in men with category III chronic prostatitis: a preliminary prospective, double-blind, placebo-controlled trial. Urology. 1999;54(6):960-963. (PubMed)

116. Ferry DR, Smith A, Malkhandi J, et al. Phase I clinical trial of the flavonoid quercetin: pharmacokinetics and evidence for in vivo tyrosine kinase inhibition. Clin Cancer Res. 1996;2(4):659-668. (PubMed)

117. Bonkovsky HL. Hepatotoxicity associated with supplements containing Chinese green tea (Camellia sinensis). Ann Intern Med. 2006;144(1):68-71. (PubMed)

118. Javaid A, Bonkovsky HL. Hepatotoxicity due to extracts of Chinese green tea (Camellia sinensis): a growing concern. J Hepatol. 2006;45(2):334-335; author reply 335-336. (PubMed)

119. Jatoi A, Ellison N, Burch PA, et al. A phase II trial of green tea in the treatment of patients with androgen independent metastatic prostate carcinoma. Cancer. 2003;97(6):1442-1446. (PubMed)

120. Pisters KM, Newman RA, Coldman B, et al. Phase I trial of oral green tea extract in adult patients with solid tumors. J Clin Oncol. 2001;19(6):1830-1838. (PubMed)

121. Chow HH, Cai Y, Hakim IA, et al. Pharmacokinetics and safety of green tea polyphenols after multiple-dose administration of epigallocatechin gallate and polyphenon E in healthy individuals. Clin Cancer Res. 2003;9(9):3312-3319. (PubMed)

122. Bailey DG, Dresser GK. Interactions between grapefruit juice and cardiovascular drugs. Am J Cardiovasc Drugs. 2004;4(5):281-297. (PubMed)

123. Dahan A, Altman H. Food-drug interaction: grapefruit juice augments drug bioavailability--mechanism, extent and relevance. Eur J Clin Nutr. 2004;58(1):1-9. (PubMed)

124. Zhou S, Lim LY, Chowbay B. Herbal modulation of P-glycoprotein. Drug Metab Rev. 2004;36(1):57-104. (PubMed)

125. Marzolini C, Paus E, Buclin T, Kim RB. Polymorphisms in human MDR1 (P-glycoprotein): recent advances and clinical relevance. Clin Pharmacol Ther. 2004;75(1):13-33. (PubMed)

126. Hurrell RF, Reddy M, Cook JD. Inhibition of non-haem iron absorption in man by polyphenolic-containing beverages. Br J Nutr. 1999;81(4):289-295. (PubMed)

127. Zijp IM, Korver O, Tijburg LB. Effect of tea and other dietary factors on iron absorption. Crit Rev Food Sci Nutr. 2000;40(5):371-398. (PubMed)

128. Song J, Kwon O, Chen S, et al. Flavonoid inhibition of sodium-dependent vitamin C transporter 1 (SVCT1) and glucose transporter isoform 2 (GLUT2), intestinal transporters for vitamin C and Glucose. J Biol Chem. 2002;277(18):15252-15260. (PubMed)

129. US Department of Agriculture. USDA database for the flavonoid content of selected foods.

Flavonoids | Linus Pauling Institute | Oregon State University

Available at: http://www.nal.usda.gov/fnic/foodcomp/Data/Flav/flav.html. Accessed 9/13/04.

130. US Department of Agriculture. USDA database for the proanthocyanidin content of selected foods. Available at: <u>http://www.nal.usda.gov/fnic/foodcomp/Data/PA/PA.html</u>. Accessed 9/13/04.

131. Vrhovsek U, Rigo A, Tonon D, Mattivi F. Quantitation of polyphenols in different apple varieties. J Agric Food Chem. 2004;52(21):6532-6538. (PubMed)

132. Moyer RA, Hummer KE, Finn CE, Frei B, Wrolstad RE. Anthocyanins, phenolics, and antioxidant capacity in diverse small fruit: vaccinium, rubus, and ribes. J Agric Food Chem. 2002;50(3):519-525. (PubMed)

133. Lee HS. Characterization of major anthocyanins and the color of red-fleshed Budd Blood orange (Citrus sinensis). J Agric Food Chem. 2002;50(5):1243-1246. (PubMed)

134. Ryan JM, Revilla E. Anthocyanin composition of Cabernet Sauvignon and Tempranillo grapes at different stages of ripening. J Agric Food Chem. 2003;51(11):3372-3378. (PubMed)

135. Henning SM, Fajardo-Lira C, Lee HW, Youssefian AA, Go VL, Heber D. Catechin content of 18 teas and a green tea extract supplement correlates with the antioxidant capacity. Nutr Cancer. 2003;45(2):226-235. (PubMed)

#### Printer-friendly version

#### Disclaimer

The Linus Pauling Institute Micronutrient Information Center provides scientific information on the health aspects of dietary factors and supplements, food, and beverages for the general public. The information is made available with the understanding that the author and publisher are not providing medical, psychological, or nutritional counseling services on this site. The information should not be used in place of a consultation with a competent health care or nutrition professional.

The information on dietary factors and supplements, food, and beverages contained on this website does not cover all possible uses, actions, precautions, side effects, and interactions. It is not intended as nutritional or medical advice for individual problems. Liability for individual actions or omissions based upon the contents of this site is expressly disclaimed.

You may not copy, modify, distribute, display, transmit, perform, publish or sell any of the copyrightable material on this website. You may hyperlink to this website but must include the following statement:

"This link leads to a website provided by the Linus Pauling Institute at Oregon State University. [Your name] is not affiliated or endorsed by the Linus Pauling Institute or Oregon State University."





A Message from the LPI Director

### Vitamins

Minerals

Other Nutrients

**Dietary Factors** 

L-Carnitine

Coenzyme Q10

Lipoic Acid

Phytochemicals

Carotenoids

Chlorophyll and Chlorophyllin

Curcumin

Fiber

Flavonoids

Garlic

Indole-3-Carbinol

Isothiocyanates

Lignans

Phytosterols

Resveratrol

Soy Isoflavones

Food and Beverages

Micronutrients and Health

Flavonoids | Linus Pauling Institute | Oregon State University

Life Stages



Receive the Free LPI Newsletter



Micronutrients for Health Handout

f

**>>** 

1

Like us on Facebook

Follow us on Twitter

Get the latest from the LPI Blog

307 Linus Pauling Science Center Corvallis, OR 97331 Send Email

#### Contact Info

Linus Pauling Institute | Oregon State University 307 Linus Pauling Science Center | Corvallis, Oregon 97331 phone: 541-737-5075 | fax: 541-737-5077 email: lpi@oregonstate.edu

http://lpi.oregonstate.edu/mic/dietary-factors/phytochemicals/flavonoids[2016/01/28 21:36:31]

<u>Copyright</u> ©2016 Oregon State University <u>Disclaimer</u>